

Original article

# Kuei-Lu-Er-Xian-Jiao extract enhances BMP-2 production in osteoblasts

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## ABSTRACT

Osteoporosis is a common skeletal disorder, resulting from an imbalance in bone resorption relative to formation. Bone morphogenetic protein (BMP) is a key regulator in bone formation and osteoblastic differentiation. Hence, compounds that promote BMP expression may be suitable candidates for osteoporosis treatment. This study examined the effects of the traditional Chinese medicinal agent, Kuei-Lu-Er-Xian-Jiao (KLEXJ), on BMP-2 production in osteoblasts. We found that KLEXJ extract promoted osteoblastic differentiation marker ALP activity and increased BMP-2 production; pretreatment with PI3K and Akt inhibitors, or small interfering RNA (siRNA), reduced these effects. KLEXJ also enhanced PI3K and Akt activity, and siRNA-mediated KLEXJ-enhancement of ALP activity and BMP-2 production. KLEXJ also significantly promoted p65 phosphorylation, while treatment with PI3K and Akt inhibitors antagonized KLEXJ-enhanced p65 phosphorylation. Thus, KLEXJ enhances ALP activity and BMP-2 production of osteoblasts through the PI3K/Akt pathway.

## 1. Introduction

Bone is a mineralized organ containing several types of cells, including osteoblasts (bone-forming cells) and osteoclasts (bone-resorbing cells), which subject bone to a continuous renewal and repair process during the life of each individual by the process of bone remodelling [1, 2]. Bone resorption and osteogenic functions must be in balance in order to maintain a constant bone mass [3, 4]. Compounds that promote osteoblastic proliferation or enhance differentiation of osteoblasts result in increased bone formation [5-7]. At this time, teriparatide, the recombinant 1-34 fragment of human parathyroid hormone (rhPTH1-34), is the first bone formation agent to be approved for the treatment of osteoporosis [8, 9]. Up until now, the detailed molecular mechanism of osteoporosis has remained unclear, albeit the process is probably correlated with reduced availability or activity of growth factors. For example, bone morphogenetic proteins (BMPs), [10] were first discovered due to their ability to promote bone formation in rodents. The protein structure of BMPs resembles that of the

Abbreviations: ALP, alkaline phosphatase; BMP, bone morphogenetic protein; KLEXJ, Kuei-Lu-Er-Xian-Jiao; TCM, Traditional Chinese Medicine; FBS, fetal bovine serum; qPCR, quantitative real-time polymerase chain reaction.

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WUDQVIRUPLQJ JURZWK IDFRU VX 2.1. ALP activity assay@ , W LV NQRZQ WKDW BMP-2 plays a critical role in osteoblastic differentiation and bone

formation by increasing osteopontin, collagen and proteoglycan production, as well as promoting alkaline phosphatase (ALP) activity [12]. Previous research has also linked osteoporosis with BMP-2 gene, confirming an association with osteoporosis [13].

Traditional Chinese Medicine (TCM) is a popular component of health care in Taiwan that provides one therapeutic option for osteoporosis treatment. Emerging studies indicate that TCM promotes bone formation and prevents bone loss in the ovariectomized rat model [14, 15]. The TCM drug Kuei-Lu-Er-Xian-Jiao (KLEXJ) is a multicomponent Chinese herbal supplement that has been used for treatment of degenerative joint diseases with adverse effects for over 2,000 years [16, 17]. However, its role in osteoblastic function remains largely unknown. We report that KLEXJ extract increases osteoblastic differentiation marker ALP activity and BMP-2 production in osteoblasts, while simultaneously inhibits osteoblasts, while simultaneously visualized by enhanced chemiluminescence, using a Fujifilm R X V O \ P H G L D W L Q J W K H 3 , . \$ N W 1 ) % L A S E R 3 0 0 0 Z e n i l u m P r e s e n c e D e t e c t i o n S y s t e m (Fujifilm; Tokyo, Japan).

## 2. Experimental section

### 2.1. Materials

Kuei-Lu-Er-Xian-Jiao (KLEXJ) contains Testudinis Plastrum (species: *Chinemys reevesii*; animal part: plastrum); Cervi cornu (species: *Cervus elaphus*; animal part: antler); Radix Ginseng (species: *Panax ginseng* C. A. Meyer; plant part: root) and Lycii fructus (species: *Lycium barbarum*; plant part: fruit) and was prepared as follows: Testudinis Plastrum and Cervi cornu were stewed for 7 days, after which Radix Ginseng and Lycii fructus were added into the mixture. A 6.25 g extract was derived from the ratio between the 4 components, consisting of about 1.25 g of Testudinis Plastrum, 10 g of Cornu cervi, 0.55 g of Radix Ginseng, 1.1 g of Lycii fructus, which was provided by the Li-

An Biotechnology Pharmaceutical Company (Tainan; Taiwan). Li-An Biotechnology Pharmaceutical Company was awarded the Good Manufacturing Practice certification in Taiwan (Drug license number-013857, issued by the Department of Health, Taiwan).

Rabbit monoclonal antibodies specific for BMP-2, p85, Akt, p65, p-p85, p-Akt, p-p65 and b-actin, as well as anti-mouse anti-rabbit IgG-conjugated horseradish peroxidase, were all purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). The BMP-2 ELISA kit was obtained from Biosource Technology (Nivelles, Belgium). TRIzol reagent, Lipofectamine 2000 and the MMLV RT kit were obtained from Invitrogen (Carlsbad, CA, USA). The control, p85 and Akt siRNA were obtained from Dharmacon Research (Lafayette, CO, USA). The TaqMan assay kit was obtained from Thermo Fisher Scientific (Grand Island, NY, USA). LY294002 and other pharmacological inhibitors were purchased from Sigma-Aldrich (St. Louis, MO, USA).

### 2.2. Cell culture

The mouse osteoblast cell line MC3T3-E1 was obtained from American Type Culture Collection (Manassas, VA, USA). Cells were maintained in humidified air containing 5% CO<sub>2</sub> at 37°C with a minimum essential medium (MEM), 10% fetal bovine serum (FBS), 100 units/ml penicillin and 100 mg/ml streptomycin (Gibco-BRL Life technologies; Grand Island, NY, USA).

### 2.5. Quantitative real-time polymerase chain reaction (qPCR)

Total RNA was extracted from MC3T3-E1 cells using TRIzol reagent. Messenger RNA was reversely transcribed to complementary DNA using an MMLV RT kit, and qPCR was then performed using the Taqman assay kit [21].

### 2.6. Statistical analysis

Data are presented as mean  $\pm$  standard error of mean (SEM). Statistical analysis of both samples used the Student's *t*-test. Statistical comparisons of more than two groups were performed by one-way analysis of variance with Bonferroni's post-hoc test; *p* < 0.05 was considered significant.

## 3. Results

### 3.1. KLEXJ enhances ALP activity and BMP-2 production in osteoblasts

Differentiated osteoblasts express high ALP activity, rendering ALP activity a key marker for osteoblastic formation [22, 23]. When we examined the role of KLEXJ in ALP activity, we found that incubation of osteoblasts with KLEXJ significantly augmented ALP activity (Fig. 1A). As BMP-2 has been reported to play a key role in osteoblastic differentiation [10], we next examined whether KLEXJ promotes osteoblastic differentiation by regulating BMP-2 expression. We found that incubation with KLEXJ stimulated BMP-2 mRNA expression and protein production, in a concentration-dependent manner (Fig. 1B&C). These combined findings indicate that KLEXJ promotes osteoblastic differentiation marker ALP activity and BMP-2 production in osteoblasts.

### 3.1. KLEXJ enhances ALP activity and BMP-2 production through the PI3K/Akt signaling pathway

PI3K activation has been described as mediating bone formation and differentiation [24, 25]. The effects of KLEXJ were tested on the PI3K pathway. Incubation with the PI3K inhibitor Ly294002 or transfection with PI3K siRNA markedly abolished KLEXJ-

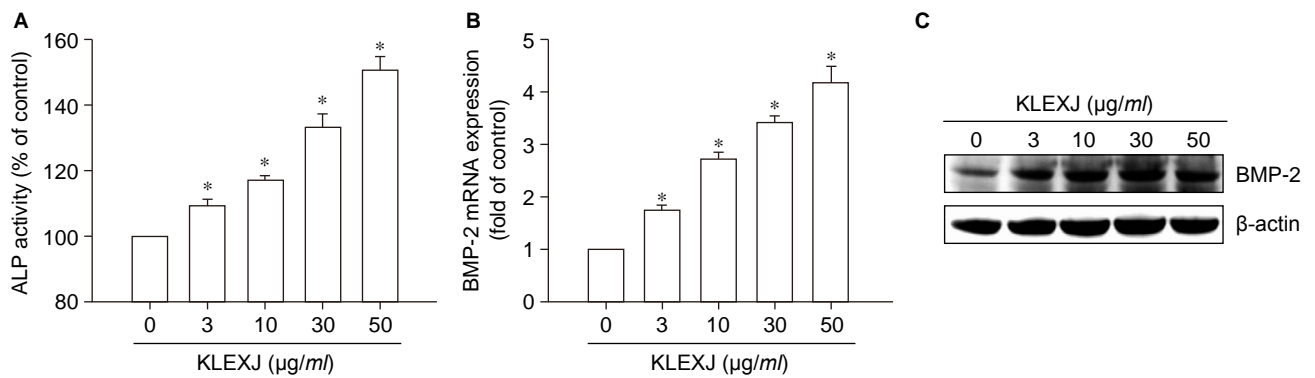


Fig. 1 - KLEXJ extract enhances ALP activity and BMP-2 expression in osteoblasts. (A) Osteoblasts were treated with KLEXJ for 48 h and ALP activity was examined by a commercial ALP assay kit. (B&C) Osteoblasts were treated with KLEXJ for 24 h and BMP-2 expression was examined by qPCR and Western blot analysis. Results are expressed as mean  $\pm$  S.E.M.,  $p < 0.05$  compared with control.

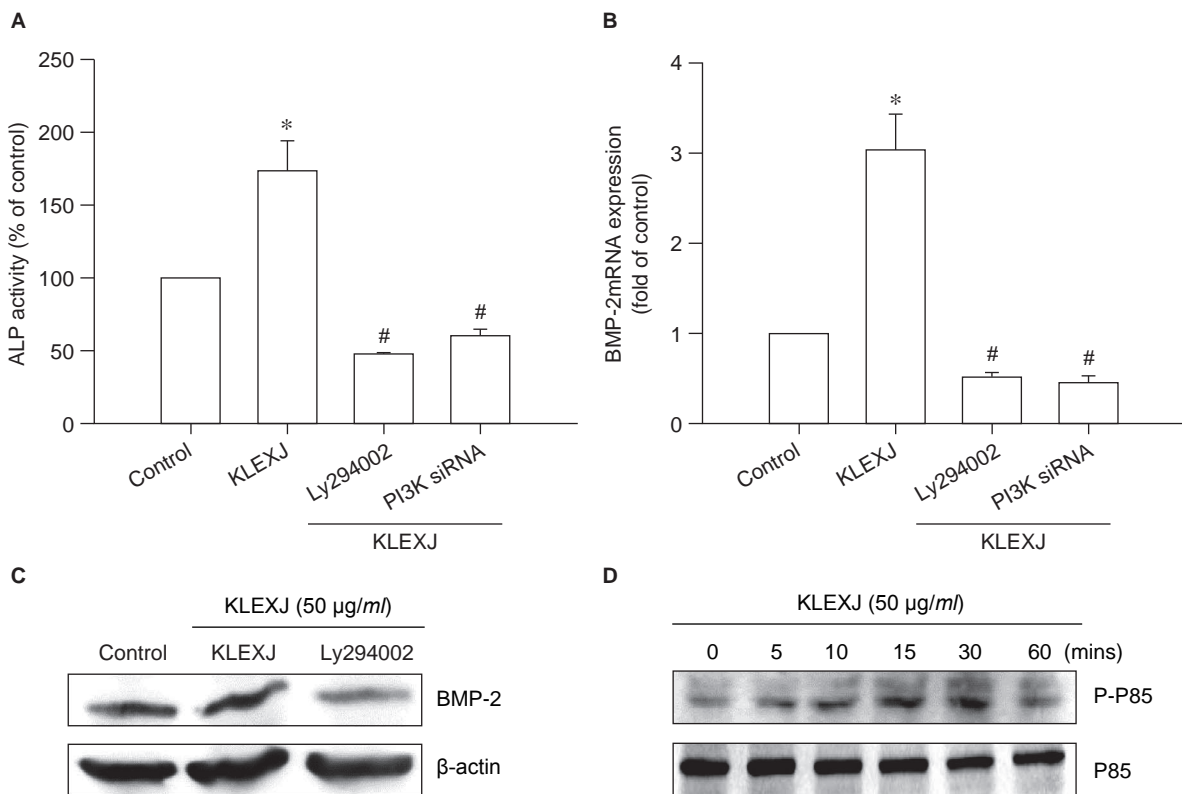


Fig. 2 - KLEXJ extract promotes ALP activity and BMP-2 expression through the PI3K pathway. (A) Osteoblasts were pretreated with Ly294002 (10  $\mu$ M) for 30 min or transfected with p85 siRNA for 24 h, followed by stimulation with KLEXJ for 48 h; ALP activity was examined with a commercial ALP assay kit. (B&C) Osteoblasts were pretreated with Ly294002 (10  $\mu$ M) for 30 min or transfected with p85 siRNA for 24 h, followed by stimulation with KLEXJ for 24 h; BMP-2 expression was examined by qPCR and Western blot analysis. (D) Osteoblasts were incubated with KLEXJ for indicated time intervals and p85 phosphorylation was examined by Western blot analysis. Results are expressed as mean  $\pm$  S.E.M.,  $p < 0.05$  compared with control. #,  $p < 0.05$  compared with KLEXJ-treated group.

induced ALP activity and BMP-2 expression (Fig. 2A-C). Treat an important role in osteoblastic function [14, 26]. We therefore ment of osteoblasts with KLEXJ promoted phosphorylation o studied whether KLEXJ also activates the Akt signaling pathway. PI3K in a time-dependent manner (Fig. 2D). Thus, KLEXJ en We found that an Akt inhibitor or Akt siRNA abolished KLEXJ- hances BMP-2 production in osteoblasts PI3K activation. induced ALP activity and BMP-2 production (Fig. 3A-C). In ad- Akt is a downstream pathway in PI3K signaling and plays dition, Akt phosphorylation was increased after KLEXJ stimula

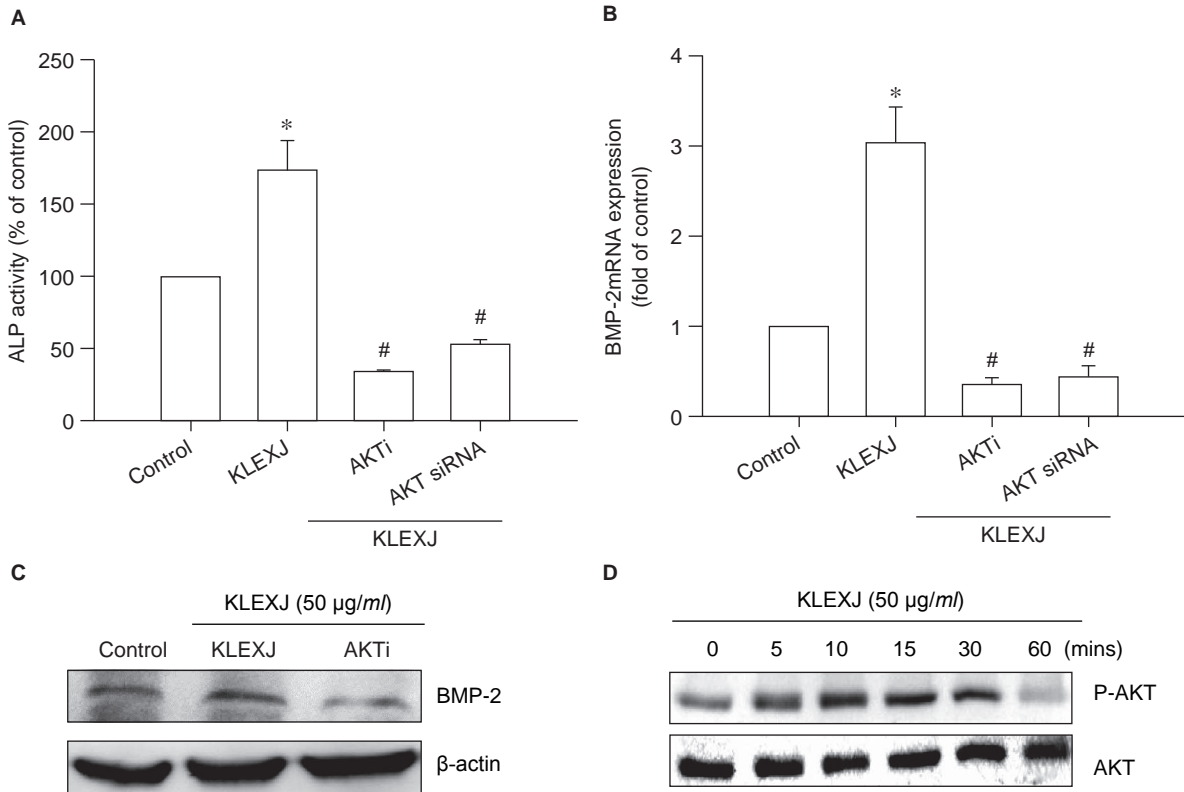


Fig. 3 - KLEXJ extract promotes ALP activity and BMP-2 expression through the Akt pathway. (A) Osteoblasts were pretreated with an Akt inhibitor (10  $\mu$ M) for 30 min or transfected with Akt siRNA for 24 h, followed by stimulation with KLEXJ for 48 h. ALP activity was examined using a commercial ALP assay kit. (B&C) Osteoblasts were pretreated with an Akt inhibitor (10  $\mu$ M) for 30 min or transfected with Akt siRNA for 24 h, followed by stimulation with KLEXJ for 24 h. BMP-2 expression was examined by qPCR and Western blot analysis. (D) Osteoblasts were incubated with KLEXJ for indicated time intervals and Akt phosphorylation was examined by Western blot analysis. Results are expressed as mean  $\pm$  S.E.M. \*,  $p < 0.05$  compared with control. #,  $p < 0.05$  compared with KLEXJ-treated group.

tion (Fig. 3D), suggesting that KLEXJ enhances ALP activity and BMP-2 production in osteoblasts through the Akt pathway.

### 3.2. KLEXJ increases ALP activity and BMP-2 production in osteoblasts via the NF- $\kappa$ B pathway

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KLEXJ extract induces ALP activity (an important osteoblastic differentiation marker). In addition, we suggest that BMP-2 acts as a target molecule of KLEXJ-induced signaling that requires the

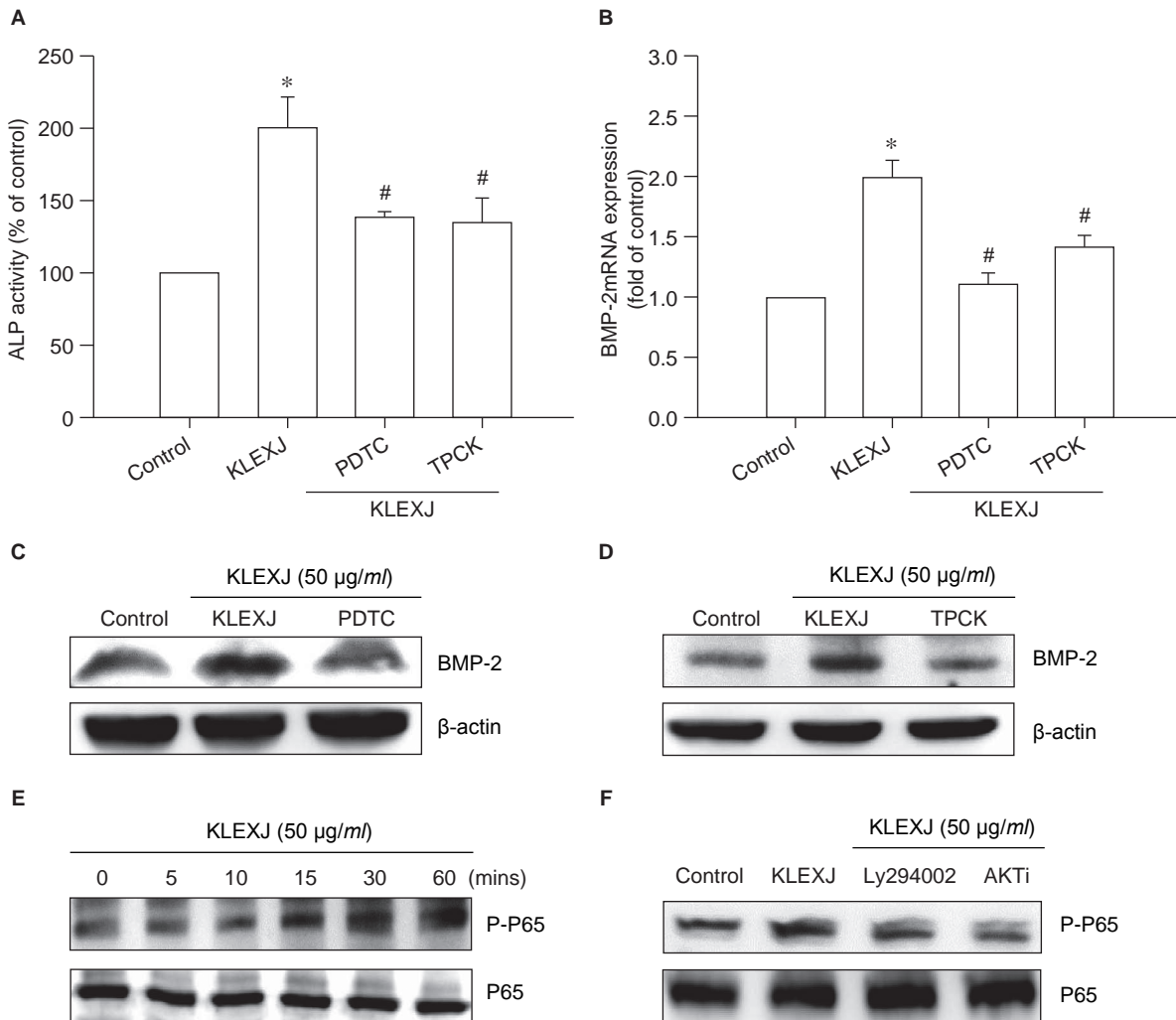
The detailed molecular mechanisms of osteoporosis remain unknown, but they are believed to correlate with decreased availability of bone growth factors, BMPs, and their receptors. BMP-2 is a key factor in bone remodeling and formation [29, 30]. The present study shows that KLEXJ extract increases production of BMP-2. In contrast, incubation of osteoblasts with KLEXJ increased and PDTC reversed KLEXJ-induced ALP activity and increases osteoblastic differentiation marker expression (ALP activity). Furthermore, KLEXJ in-creased phosphorylation of p65 in a time-dependent manner (Fig. 4E). In contrast, KLEXJ-induced activation of p65 was antagonized by pretreatment with Ly294002 and an Akt inhibitor (Fig. 4F) and a PI3K inhibitor (Fig. 4G).

PI3K activation is a potential signaling pathway that regulates bone formation [31, 32]. Here, we report that both a PI3K inhibitor and a RNF4 antagonist KLEXJ promoted activity of ALP and production of BMP-2. Incubation of osteoblasts with KLEXJ increased phosphorylation of PI3K, suggesting that PI3K activation plays a crucial role in KLEXJ-increased bone differentiation and BMP-2 expression.

Akt activation is reportedly mediates ALP activity during osteoblastic cell differentiation [33, 34]. In this study, we found that KLEXJ promotes phosphorylation of Akt, while an Akt inhibitor or siRNA diminishes KLEXJ-induced potentiation of ALP activity and BMP-2 production in osteoblasts, which suggests that Akt activation plays a critical role in KLEXJ-promoted osteoblastic

## 4. Discussion

Kuei-Lu-Er-Xian-Jiao (KLEXJ), a TCM formula, is widely used in traditional medicine for osteoporosis treatment and has been reported to reduce osteoarthritis progress [16]. However, the detailed effects of KLEXJ in bone cells are unclear. To the best of our knowledge, this study is the first analysis of the role played by KLEXJ extract in osteoblasts. Our results demonstrate that



**Fig. 4 - KLEXJ promotes ALP activity and BMP-2 expression through the NF- $\kappa$ B pathway.** (A) Osteoblasts were pretreated with TPCK (10  $\mu$ M) or PDTC (10  $\mu$ M) for 30 min, followed by stimulation with KLEXJ for 48 h. ALP activity was examined using a commercial ALP assay kit. (B-D) Osteoblasts were pretreated with TPCK (10  $\mu$ M) or PDTC (10  $\mu$ M) for 30 min, followed by stimulation with KLEXJ for 24 h and BMP-2 expression was examined by qPCR and Western blot analysis. (E) Osteoblasts were incubated with KLEXJ for indicated time intervals and p65 phosphorylation was examined by Western blot analysis. (F) Osteoblasts were pretreated with Ly294002 or an Akt inhibitor, followed by stimulation with KLEXJ. p65 phosphorylation was examined by Western blot analysis. Results are expressed as mean  $\pm$  S.E.M. \* $p$  < 0.05 compared with control. # $p$  < 0.05 compared with KLEXJ-treated group.

function.

The BMP-2 5' promoter region contains the ERE, AP-1 and Sp1 binding sites, which regulate BMP-2 expression [35]. In this study, we have demonstrated that KLEXJ has a key role in osteoblasts. In addition, treatment with KLEXJ-pretreated cells increases ALP activity and BMP-2 production. Further evidence that activation of the NF- $\kappa$ B pathway may be suitable in the treatment of osteoporosis.

Conclusions  
 Osteoblasts have a key role in bone formation and bone maintenance. KLEXJ has been shown to be a key role in osteoblasts. In addition, treatment with KLEXJ-pretreated cells increases ALP activity and BMP-2 production. Further evidence that activation of the NF- $\kappa$ B pathway may be suitable in the treatment of osteoporosis.

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## Conflict of interest

None of the authors have any financial or personal relationships with other people or organizations that could inappropriately influence this work.

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## REFERENCES

- [1] Manolagas SC, Jilka RL. Bone marrow, cytokines, and bone remodeling. Emerging insights into the pathophysiology of osteoporosis. *N Engl J Med.* 1995; 332: 305-11.
- [2] Chen PC, Cheng HC, Yang SF, Lin CW, Tang CH. The CCN family proteins: modulators of bone development and novel targets in bone-associated tumors. *Biomed Res Int.* 2014; 2014: 437096.
- [3] Goltzman D. Discoveries, drugs and skeletal disorders. *Nat Rev Drug Discov.* 2002; 1: 784-96.
- [4] Wong SK, Chin KY, Suhaimi FH, Ahmad F, Ima-Nirwana S. The Relationship between Metabolic Syndrome and Osteoporosis: A Review. *Nutrients.* 2016; 8.
- [5] Ducy P, Schinke T, Karsenty G. The osteoblast: a sophisticated fibroblast under central surveillance. *Science.* 2000; 289: 1501-4.
- [6] Lane NE, Kelman A. A review of anabolic therapies for osteoporosis. *Arthritis Res Ther.* 2003; 5: 214-22.
- [7] Wu JB, Fong YC, Tsai HY, Chen YF, Tsuzuki M, Tang CH. -Narigin-induced bone morphogenetic protein-2 expression via PI3K, Akt, c-Fos/c-Jun and AP-1 pathway in osteoblasts. *Eur J Pharmacol.* 2008; 588: 333-41.
- [8] Fox J. Developments in parathyroid hormone and related peptides as bone-formation agents. *Curr Opin Pharmacol.* 2002; 2: 338-44.
- [9] Lindsay R, Krege JH, Marin F, Jin L, Stepan JJ. Teriparatide for osteoporosis: importance of the full course. *Osteoporos Int.* 2016; 27: 2395-410.
- [10] Rider CC, Mulloy B. Bone morphogenetic protein and growth differentiation factor cytokine families and their protein antagonists. *Biochem J.* 2010; 429: 1-12.
- [11] Thatcher JD. The TGF-beta signal transduction pathway. *Sci Signal.* 2010; 3: tr4.
- [12] Hong CC, Yu PB. Applications of small molecule BMP inhibitors in physiology and disease. *Cytokine Growth Factor Rev.* 2009; 20: 409-18.
- [13] Styrkarsdottir U, Cazier JB, Kong A, Rolfsson O, Larsen H, Bjarnadottir E, et al. Linkage of osteoporosis to chromosome 20p12 and association to BMP2. *PLoS Biol.* 2003; 1: E69.
- [14] Wu CM, Chen PC, Li TM, Fong YC, Tang CH. Si-Wu-tang extract stimulates bone formation through PI3K/Akt/NF-kappaB signaling pathways in osteoblasts. *BMC Complement Altern Med.* 2013; 13: 277.
- [15] Li TM, Huang HC, Su CM, Ho TY, Wu CM, Chen WC, et al. Cistanche deserticola extract increases bone formation in osteoblasts. *J Pharm Pharmacol.* 2012; 64: 897-907.
- [16] Tsai CC, Chou YY, Chen YM, Tang YJ, Ho HC, Chen DY. Effect of the herbal drug guilu erxian jiao on muscle strength, articular pain, and disability in elderly men with knee osteoarthritis. *Evid Based Complement Alternat Med.* 2014; 2014: 297458.
- [17] Lee SC, Chang SJ, Tsai LY. Effects of traditional Chinese medicines on serum lipid profiles and homocysteine in the ovariectomized rats. *Am J Chin Med.* 2004; 32: 541-50.
- [18] Tang CH, Hsu CJ, Fong YC. The CCL5/CCR5 axis promotes interleukin-6 production in human synovial fibroblasts. *Arthritis Rheum.* 2010; 62: 3615-24.
- [19] Chien SY, Huang CY, Tsai CH, Wang SW, Lin YM, Tang CH. Interleukin-1beta induces fibroblast growth factor 2 expression and subsequently promotes endothelial progenitor cell angiogenesis in chondrocytes. *Clin Sci (Lond).* 2016; 130: 667-81.
- [20] Yang WH, Chang AC, Wang SW, Wang SJ, Chang YS, Chang TM, et al. Leptin promotes VEGF-C production and induces lymphangiogenesis by suppressing miR-27b in human chondrosarcoma cells. *Sci Rep.* 2016; 6: 28647.
- [21] Huang CY, Chang AC, Chen HT, Wang SW, Lo YS, Tang CH. Adiponectin promotes VEGF-C-dependent lymphangiogenesis by inhibiting miR-27b through a CaMKII/AMPK/p38 signaling pathway in human chondrosarcoma cells. *Clin Sci (Lond).* 2016; 130: 1523-33.
- [22] Tang CH, Hsu TL, Lin WW, Lai MZ, Yang RS, Hsieh SH, et al. Attenuation of bone mass and increase of osteoclast formation in decoy receptor 3 transgenic mice. *J Biol Chem.* 2007; 282: 2346-54.
- [23] Tang CH, Yang RS, Chien MY, Chen CC, Fu WM. Enhancement of bone morphogenetic protein-2 expression and bone formation by coumarin derivatives via p38 and ERK-dependent pathway in osteoblasts. *Eur J Pharmacol.* 2008; 579: 40-9.
- [24] Li H, Li T, Fan J, Fan L, Wang S, Weng X, et al. miR-216a rescues dexamethasone suppression of osteogenesis, promotes osteoblast differentiation and enhances bone formation, by regulating c-Cbl-mediated PI3K/AKT pathway. *Cell Death Differ.* 2015; 22: 1935-45.
- [25] Su CM, Lee WL, Hsu CJ, Lu TT, Wang LH, Xu GL, et al. Adiponectin Induces Oncostatin M Expression in Osteoblasts through the PI3K/Akt Signaling Pathway. *Int J Mol Sci.* 2016; 17.
- [26] Chen HT, Tsou HK, Chang CH, Tang CH. Hepatocyte growth factor increases osteopontin expression in human osteoblasts through PI3K, Akt, c-Src, and AP-1 signaling pathway. *PLoS One.* 2012; 7: e38378.
- [27] Tan TW, Huang YL, Chang JT, Lin JJ, Fong YC, Kuo CC, et al. CCN3 increases BMP-4 expression and bone mineralization in osteoblasts. *J Cell Physiol.* 2012; 227: 2531-41.
- [28] Kageyama A, Matsui H, Ohta M, Sambuichi K, Kawano H, Notsu T, et al. Palmitic acid induces osteoblastic differentiation in vascular smooth muscle cells through ACSL3 and NF-kappaB, novel targets of eicosapentaenoic acid. *PLoS One.* 2013; 8: e68197.
- [29] Wang J, Guo J, Liu J, Wei L, Wu G. BMP-Functionalised Coatings to Promote Osteogenesis for Orthopaedic Implants. *Int J Mol Sci.* 2014; 15: 10150-68.
- [30] Bandyopadhyay A, Yadav PS, Prashar P. BMP signaling in development and diseases: a pharmacological perspective. *Biochem Pharmacol.* 2013; 85: 857-64.
- [31] Chen CY, Su CM, Huang YL, Tsai CH, Fuh LJ, Tang CH. CCN1 in-

- duces oncostatin M production in osteoblasts integrin-dependent signal pathways. *PLoS One*. 2014; 9: e106632.
- [32] Pramojane SN, Phimphilai M, Chattipakorn N, Chattipakorn SC. Possible roles of insulin signaling in osteoblasts. *Endocr Res*. 2014; 39: 144-51.
- [33] Lai CH, Wu YW, Yeh SD, Lin YH, Tsai YH. Effects of 6-Hydroxyflavone on Osteoblast Differentiation in MC3T3-E1 Cells. *Evid Based Complement Alternat Med*. 2014; 2014: 924560.
- [34] Marie PJ. Signaling pathways affecting skeletal health. *Curr Osteoporos Rep*. 2012; 10: 190-8.
- [35] Hou CH, Hou SM, Tang CH. Ultrasound increased BMP-2 expression via PI3K, Akt, c-Fos/c-Jun, and AP-1 pathways in cultured osteoblasts. *J Cell Biochem*. 2009; 106: 7-15.
- [36] Yin MC. Development of natural antitumor agents. *BioMedicine*. 2013; 3: 105.
- [37] Su KP. Nutrition, psychoneuroimmunology and depression: the therapeutic implications of omega-3 fatty acids in interferon-alpha-induced depression. *BioMedicine*. 2015; 5: 21.
- [38] Yao HT, Yang YC, Chang CH, Yang HT, Yin MC. Protective effects of (-)-epigallocatechin-3-gallate against acetaminophen-induced liver injury in rats). *BioMedicine*. 2015; 5: 15.
- [39] Zhai YK, Guo X, Pan YL, Niu YB, Li CR, Wu XL et al. A systematic review of the efficacy and pharmacological profile of *Herba Epimedii* in osteoporosis therapy. *Pharmazie*. 2013; 68: 713-22.
- [40] Keiler AM, Zierau O, Kretzschmar G. Hop extracts and hop substances in treatment of menopausal complaints. *Planta Med*. 2013; 79: 576-9.